

Racemization Hypothesis of COVID-19. Tip of the Iceberg

Journal of Psychology and Neuroscience

Review Article

Victor V. Dyakin^{1*}, Henry Sershen¹, Nika V. Dyakina-Fagnano², Pamela D. Butler¹ and Thomas M. Wisniewski³

¹The Nathan Kline Institute for Psychiatric Research, USA

²Child, Adolescent and Young Adult Psychiatry, USA

³New York University School of Medicine, USA

*Correspondence author

Victor Vasilyevich Dyakin

PhD

The Nathan S. Kline Institute for Psychiatric Research (NKI).

Perception of Virtual Reality Lab. (PVRL).

Address: 140 Old Orangeburg Road, Bldg. 35.

Orangeburg, NY. 10962-1167. USA. Bld.35. Rom 201-C.

Phone: 845-548-96-94. Fax: (845) 398-5510.

Submitted : 16 July 2020 ; Published : 4 Aug 2020

Abstract

The impact of viral infections on the central nervous system is widely known. Virus-related neuropsychiatric and neurobehavioral syndromes are caused by the distortion of cognitive, affective, behavioral, and perceptual domains. Although it is a commonly known phenomenon, the mechanism behind it is not well-understood. The contagious and deadly features of coronavirus disease 2019 (COVID-19) have been associated with the virus-host cell interaction at the molecular level. However, there is no reliable biomarker characterizing the disease progression. Studies of the structure, function, and evolution of coronavirus transmembrane spike glycoproteins (S-, N-, and E-proteins) suggest an essential role of protein chirality in virus-cell membrane interaction. The virus-host interaction is the subject of multidisciplinary research from the biochirality and systems biology, to cell physiology and non-equilibrium thermodynamics of phase transitions in proteins. At the protein level, virus-host interaction is modulated by the amino acid sequence of viral proteins and cellular metabolism. Enzymatic and spontaneous post-translational modifications (PTMs) are two mutually influential mechanisms governing the dynamics of virus and host cell proteome. Among them, phosphorylation and racemization are the most inter-related and studied. The spontaneous phase transitions within viral glycoprotein impacts the cell-entry capability of the virus. The spontaneous racemization is a particular and highly specific metabolic event in virus-cell interaction that is the focus of our attention. Many viral proteins are characterized by a high proportion of the serine (Ser) residues, which are the common target of the host-cell glycosylation, phosphorylation, and racemization, and proteolytic enzymes. Particularly, coronavirus N proteins were found to be phosphorylated at multiple Ser residues, a portion of which are shown to be phosphorylation-prone by the Ser-associated kinases. Since Ser is known as one of the most racemization prone amino acids, we promote an idea of the specific impact of spontaneous racemization at Ser residues on virus-host interaction.

Keywords : Virus-host Cell Interaction, Virus Proteins, Post-translational Modification, Spontaneous Racemization, Spontaneous Phase Transitions

Abbreviations

Amino acids	:	AAs	Herpes simplex virus type 1	:	HSV-1
D-amino acids	:	D-AAs	Human immunodeficiency virus	:	HIV
Carboxyl-terminal domain	:	CTD	Nucleocapsid protein	:	N protein
Central nervous system	:	CNS	Post-translational modifications	:	PTMs
Coronavirus	:	CoV	Serine	:	Ser
Coronavirus disease 2019	:	COVID-19	D-Serine	:	D-Ser
			D-Aspartate	:	D-Asp

Threonine	:	Thr
Tyrosine	:	Tyr
Spike glycoprotein	:	S-proteins
Phase transitions	:	PhTs
Ribonucleic acid RNA, RNA polymerase II	:	RNAP II
Severe acute respiratory syndrome coronavirus 2	:	SARS-CoV-2

Introduction

The intra-cellular protein-based metabolic network is an evolutionary conserved, but also a highly dynamic system. The main driving force of the network is a complex of canonical enzyme-catalyzed post-translational modifications (PTMs). The number of molecular side products and environmental stresses gives rise to many spontaneous non-canonical pathways of PTMs, that are involved in protein aging and human age-related disorders [1,2,3]. Enzymatic and spontaneous PTMs are two mutually influential mechanisms. For example, excessive phosphorylation of TAU is known as a cause of protein aggregation [4,5,6]. The mechanism is believed to be cell, protein, and residue specific. Post-translational phosphoproteins display significant age-related changes in the composition of amino acid's (AA's), and in their cross-linking, and racemization. A relevant example is the age-related loss of phosphoserine (SerPh) content in human phosphoproteins. This mechanism remains to be studied and presumably is associated with the interplay of enzymatic phosphorylation and spontaneous racemization [6]. The widespread role of non-enzymatic reactions in cell metabolism is also well documented [7]. In particular, it has been shown that the spontaneous modification of AAs in viral glycoprotein impacts the cell-entry capability of the virus [8]. The significance of biochirality is supported by a shared recognition that the spontaneous or induced mutations in viral genetic material may alter the disease's pathogenesis. This spontaneous racemization, a particular and highly specific metabolic events in virus-cell interaction, is the focus of the current short review.

Biochirality of Coronavirus

The origin, transmission, and clinical therapies of coronavirus disease 2019 (COVID-19) are a primary target of current medical and scientific investigation [1-30]. Coronavirus (CoV) is a group of enveloped RNA viruses causing respiratory diseases in both humans and animals. Many details of CoV-associated damage at the cellular and molecular levels are still unclear. Due to their inability to self-replicate, viruses have developed unique potentials to utilize and modify the metabolic and signaling pathways of the host cell. Recent development shows that the host cell entry and the replication cycle of coronavirus (CoV) employs a variety of forms of the cell's protein's PTMs including glycosylation and phosphorylation [24-28]. The fundamental physical mechanism underlying the translocation of viral genomes into the cells is traditionally identified as non-equilibrium phase transitions [31-36]. However, none of them gives attention to the essential role of the chiral determinants of molecular condensation, highlighted

in several reviews [3]. The interplay of viral and host cell proteins PTMs is the subject of increasing attention. The interaction networks of viral and host proteins during early steps of infection is well known from immunodeficiency virus (HIV) related studies [29]. The knowledge about immune response induced by CoV is critical to patient treatment [37]. The mammalian innate and adaptive immune systems are under the control of AAs metabolism associated with the complex network of enzymatic and spontaneous racemization [38,39]. The pivotal role of D-amino acids (D-AAs), including D-Serine (D-Ser) and D-Aspartate (D-Asp) in the innate and adaptive immune systems, is a new-emerging and promising field of biochemistry [40-42]. However, in spite of the known role of D-AAs in immune system function, there is a void of information about the enzymatic and spontaneous racemization of CoV proteins and the impact of racemization on the virus-receptor coupling, membrane fusion, endocytosis and the host cell's protein racemization. The discovery of spontaneous PTMs in noroviruses suggests that the list of PTMs involved in this should include not only enzymatic but also spontaneous forms of PTMs including racemization and epimerization, which are common in the host cell physiology and pathology [3,30]. Notably, protein damage due to aberrant PTMs is a significant hallmark of lung aging[43]. The reductions of lung functions with age, at the molecular level, are associated with the cell type-specific protein aging and loss of functions related to aberrant PTMs. It is thus logical to assume that COVID-19 progression could be regulated by the aberrant PTMs.

The most accepted form of aberrant PTM associated with protein aging, aggregation, and dysfunction is spontaneous racemization [3]. The aberrant virus associated PTMs of proteins is well-known effect [28]. One particular example involves collagen nanofibers, which are the primary determinants of the biological and structural integrity of various tissues and organs, including bone, skin, tendon, blood vessels, cartilage, and the lungs [11]. Excessive deposition of collagen has previously been seen in virus-related pulmonary fibrosis [44]. Severe acute respiratory disease caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is associated with multi-modal lung dysfunction. At the protein level, SARS-CoV-2 is accompanied by collagen fibrosis [45]. The AA's chain of collagen contains multiple phosphorylation-prone and racemization-prone serine (Ser) residues [12-14]. The accumulation of fibrotic collagen is a well-known process that accompanies many pathological conditions [15-17]. The experimental results of lung proteomics in rodents revealed that collagen protein was increasingly racemized with age. Collagen is also a target of spontaneous racemization [18]. Most racemized AAs in lung collagen have been identified as Ser [19]. Although the details of the cellular responses to the coronavirus are unknown, the epithelial cells of the airway have been identified as a primary target [20]. Alterations in the composition of the extracellular collagen matrix have been shown in many pulmonary disorders [21,22].

Many viral proteins are characterized by a high proportion of Ser residues. Particularly, CoV nucleocapsid proteins (N

proteins) were found to be phosphorylated at multiple Ser residues, a portion of which is shown to be phosphorylation-prone by the serine-associated kinases [46-49]. Since Ser is one of the racemization prone AAs, we promote an idea concerning the specific impact of spontaneous racemization on virus-host interaction [3]. Viral Ser-targeting enzymes are traditionally used as a drug target in clinical practice [23]. This targeting strategy is in agreement with the several essential facts that point to the key role of Ser-associated enzymes in virus-host cell interactions including Ser kinases, Ser proteases, and Ser hydrolase enzymes [50-55,58].

The cell entry programs for CoV are mediated by the viral transmembrane spike glycoprotein (S protein) that bind cellular receptors and involves virus-cell membrane fusions. The interaction of S protein with the membrane receptor triggers a cascade of events including proteolysis, and acidification in endosomes [56]. It was found that S protein in viruses isolated from humans during the 2003–2004 outbreak had a Ser at position 360 (Ser-360) located in the α -helix region [57]. As a result, among the respiratory virus-activating membrane-anchored enzymes, the Ser protease family is of specific interest [58]. The function of the kinase-phosphatase PTMs at Ser, threonine (Thr), and tyrosine (Tyr) residues are well known from the studies of many human viruses including (HIV) [29,59]. Notably, Ser residues in this triad are classified as the most racemization- and phosphorylation-prone residues implicated in protein ageing and cellular disfunction [3,51]. The studies of herpes simplex virus type 1 (HSV-1) reveal that infection alters the phosphorylation of the carboxyl-terminal domain (CTD) of RNA polymerase II (RNAP II). CTD consists of a repeated heptameric sequence (YSPTSPS) containing three Ser residues (Ser-2, Ser-5, and Ser-7). Phosphorylation at Ser-2, Ser-5, and Ser-7 is essential for enzyme function and is vulnerable to the viral impact [60,61]. Protein phosphorylation on serine, threonine, and tyrosine (Ser/Thr/Tyr) is considered to be the major regulatory PTMs in eukaryotic cells from bacteria to mammal [62]. In mammals, the racemization-prone Ser residues are most closely linked to the post-translational phosphorylation (PTPh) of proteins Accordingly, Ser is among the key players in the rapid evolution of protein phosphorylation sites [3,63]. Also, PTPh of Ser serves for the rewiring and modulation of the cell signal pathways. Along with the Tyr and lysine (Lys), Ser phosphorylation is an essential regulator of NMDAR-associated neurotransmission [64]. From the broader biological scale, Ser associated phosphorylation is known as a mechanism involved in natural selection to fit the environment.

We hypothesize that the same Ser-centered mechanisms should be considered not only for the acute response to the environmental cues in general, but also for the immune response to the viral infection.

Conclusion

The coherent set of evidence discussed above, allows for the articulation of the hypothesis that COVID-19 triggers a cascade of spontaneous PTM. In this regard, the control of the

level of D-AAAs (including D-Ser) in the lung can serve as a reliable biomarker of COVID-19- related disease conditions. This hypothesis is supported by the newly derived line of facts including the finding that viral infection {herpes simplex virus type I (HSV-1)} of human-induced neural stem cells (hiNSCs) {3D bioengineered brain model} leads to the formation of the amyloid plaque-like aggregations [65]. The aggregates of amyloid plaque contain several D-AAAs. It is also known that D-AAAs-containing proteins are resistant to the metabolic and digestive enzymes, which usually recognize only proteins composed exclusively of L-AAAs-based proteins or peptides [66]. As a result, unmetabolized D-AAAs-containing brain peptides may be found in urine and blood, serving as the useful biomarker for associated diseases. The broad range significance of bio-chirality is exemplified in the studies of D-AAAs role in kidney-related and neurodegenerative diseases [2,67,68]. Further investigation of the epidemiology, pathogenesis, and chiral proteomics of the virus is necessary for the proper understanding of acute and long-term consequences of CoV infections, as well as for nutritional support to patients, and the development of effective therapeutic and prophylactic [71].

Afterword

The multidisciplinary facets of virus-related research require attention to the physics of protein folding [3]. From a biophysics perspective, the capsid shell of the virus is seen as a two-dimensional crystal with a limited size closed surface comprising inside cargo space [70].

The capsid surface is the arrangement of the equivalent oligomeric sub-units. The chirality of virus proteins is a critical internal determinant of the handedness observed in capsid morphology. The chirality propagation from the protein to the morphological level is equally necessary for two different events [71]. The first: virus entry into the host cell and the second is the protection from unnecessary molecular invasions into virus DNA [72]. The spontaneous phase transitions (PhTs) within viral glycoproteins impacts the cell-entry capability of the virus. The spontaneous racemization is a particular and highly specific metabolic event in virus-cell interaction, and is the focus of our attention. Many viral proteins are characterized by a high proportion of the Ser residues, which are the common target of the host-cell glycosylation, phosphorylation, racemization, and proteolytic enzymes [58,73-76]. Particularly, CoV N proteins were found to be phosphorylated at multiple Ser residues, a portion of which are shown to be phosphorylation-prone by the Ser-associated kinases. Since Ser is known as one of the most racemization prone amino acids, we promote the idea of the importance of the specific impact of spontaneous racemization on virus-host interaction. While speculative, the above-considered arguments are convincing to expect that spontaneous racemization may also be involved in the development of neuropsychiatric and cognitive pathologies.

Acknowledgment

Authors express acknowledgment to Abel Lajtha and Alexander G. Dadali for useful consultation.

Authors Conflict

There is NO conflict of interest to disclose.

There are no sources of funding.

References

- Wagner GR, Hirschey M.D (2014) Nonenzymatic protein acylation as a carbon stress regulated by sirtuin deacylases. *Mol. Cell* 54: 5-16. Doi: 10.1016/j.molcel.2014.03.027.
- Gabriel Piedrafitra, Markus A Keller, and Markus Ralsler (2015) The Impact of Non-Enzymatic Reactions and Enzyme Promiscuity on Cellular Metabolism during (Oxidative) Stress Conditions. *Biomolecules* 5(3): 2101–2122. Doi: 10.3390/biom5032101.
- Victor V. Dyakin, Thomas M. Wisniewski, and Abel Lajtha (2020) Chiral Interface of Amyloid Beta (A β): Relevance to Protein Aging, Aggregation and Neurodegeneration. *Symmetry* 12(4): 585. Doi.org/10.3390/sym12040585.
- Wendy Noble, Diane P Hanger, Christopher C J Miller, Simon Lovestone (2013) The Importance of Tau Phosphorylation for Neurodegenerative Diseases. *Front Neurol* 4: 83. Doi: 10.3389/fneur.2013.00083.
- Joerg Neddens, Magdalena Temmel, Stefanie Flunkert, Bianca Kerschbaumer, Christina Hoeller, Tina Loeffler, Vera Niederkofler, Guenther Daum, Johannes Attems & Birgit Hutter-Paier (2018) Phosphorylation of different tau sites during progression of Alzheimer's disease. *Acta Neuropathologica Communications* 6: 52. Article number: 52 (2018). Doi.org/10.1186/s40478-018-0557-6.
- Cloos PA, Jensen AI (2000) Age-related De-Phosphorylation of Proteins in Dentin: A Biological Tool for Assessment of Protein Age. *Biogerontology* 1(4): 341-356. Doi: 10.1023/a:1026534400435.
- Keller Mk, Piedrafitra G, Ralsler M (2015) The widespread role of non-enzymatic reactions in cellular metabolism. *Current Opinion in Biotechnology* 34: 153-161. Doi.org/10.1016/j.copbio.2014.12.020
- John B. Ruedas, Jason T. Ladner, Chelsea R. Ettinger, Suryarm Gummuru, Gustavo Palacios, John H. Connor. Lyles DS Editor (2017) Spontaneous Mutation at Amino Acid 544 of the Ebola Virus Glycoprotein Potentiates Virus Entry and Selection in Tissue Culture. *J. of Virology* 91: 15 e00392-17. Doi: 10.1128/JVI.00392-17
- Guo Y, Cao Q, Hong Z, et al. (2020) The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak – an update on the status. *Military Med. Res* 7: 1. Doi.org/10.1186/s40779-020-00240-0.
- Bianca S. Heinrich, Zoltan Maliga, David A. Stein, Anthony A. Hyman, Sean P. J. Whelan. Peter Palese, Editor (2018) Phase Transitions Drive the Formation of Vesicular Stomatitis Virus Replication Compartments. *MBio* 9: 5: e02290-17. Doi: 10.1128/mBio.02290-17.
- Caroline Hayward, Xinhua Shu, Artur V. Cideciyan, Alan Lennon, Perdita Barran, Sepideh Zarepari, Lindsay Sawyer, Grace Hendry, Baljean Dhillon, Ann H. Milam (2003) Mutation in a short-chain collagen gene, CTRP5, results in extracellular deposit formation in late-onset retinal degeneration: a genetic model for age-related macular degeneration. *Human Molecular Genetics* 12(20): 2657–2667. Doi.org/10.1093/hmg/ddg289.
- Okamoto M, Yonejima Y, Tsujimoto Y, Suzuki Y, Watanabe K (2001) A thermostable collagenolytic protease with a very large molecular mass produced by thermophilic *Bacillus* sp. strain MO-1. *Appl. Microbiol. Biotechnol* 57: 103–108.
- Zhao, GY, Chen XL, Zhao HL, Xie BB, Zhou BC, Zhang YZ (2008) Hydrolysis of insoluble collagen by diseasein MCP-01 from deep-sea *Pseudoalteromonas* sp. SM9913: collagenolytic characters, collagen-binding ability of C-terminal polycystic kidney disease domain, and implication for its novel role in deep-sea sedimentary particulate organic nitrogen degradation. *J. Biol. Chem* 283(52): 36100–36107. Doi: 10.1074/jbc.M804438200.
- Chen X, Peng M, Li J, et al. (2017) Preparation and functional evaluation of collagen oligopeptide-rich hydrolysate from fish skin with the serine collagenolytic protease from *Pseudoalteromonas* sp. SM9913. *Sci Rep* 15716. Doi.org/10.1038/s41598-017-15971-9.
- J Uitto, E M Tan, L Ryhänen (1982) Inhibition of Collagen Accumulation in Fibrotic Processes: Review of Pharmacologic Agents and New Approaches with Amino Acids and their Analogues. *J Invest Dermatol* 79(1): 113s-120s. Doi: 10.1111/1523-1747.ep12545951.
- Birk DE, and Trelstad RL (1986) Extracellular compartments in tendon morphogenesis: collagen fibril, bundle, and macroaggregate formation. *J Cell Biol* 103: 231-240.
- [Pardo et al. 2016] Pardo A, Cabrera S, Maldonado M, et al. (2016) Role of matrix metalloproteinases in the pathogenesis of idiopathic pulmonary fibrosis. *Respir Res* 17: 23. Doi.org/10.1186/s12931-016-0343-6.
- Garnero P, Cloos P, Sornay-Rendu E, Qvist P, Delmas PD (2002) Type I collagen racemization and isomerization and the risk of fracture in postmenopausal women: the OFELY prospective study. *J Bone Miner Res* 17(5): 826-833. Doi: 10.1359/jbmr.2002.17.5.826.
- Kasai A, Yamashita N, Utsunomiya-Tate N (2010) Collagen Racemization and Deposition in the Lungs of Aged Rats. *Biochemistry Insights* 3: 25–33. Doi.org/10.4137/BCI.S4210.
- Robert J, Mason RJ (2020) Pathogenesis of COVID-19 from a cell biology perspective. *European Respiratory Journal*. 2020. 55: 2000607. Doi: 10.1183/13993003.00607-2020.
- Renata Suman Mascaretti, Marta Maria Galli Bozzo Mataloun, Marisa Dolhnikoff, Celso Moura Rebello (2009) Lung Morphometry, Collagen and Elastin Content: Changes After Hyperoxic Exposure in Preterm Rabbits. *Clinics (Sao Paulo)*. 2009. 64(11): 1099-1104. Doi:

- 10.1590/S1807-59322009001100010.
22. Mereness JA, Bhattacharya S, Wang Q, Ren Y, Pryhuber GS, Mariani TJ (2018) Type VI collagen promotes lung epithelial cell spreading and wound-closure. *PLOS ONE*. 2018. 3: 12:e0209095. Doi.org/10.1371/journal.pone.020909.
 23. Marcin Skoreński, Renata Grzywa, Marcin Sieńczyk (2016) Why should we target viral serine proteases when developing antiviral agents? *Future Virology*. 2016. 11: 12. Doi.org/10.2217/fvl-2016-0106.
 24. de Wit E, van Doremalen N, Falzarano D (2016) Munster VJ. SARS and MERS: recent insights into emerging coronaviruses. *Nat. Rev. Micro* 14: 523. Doi: 10.1038/nrmicro.2016.81.
 25. Poppe, M. Wittig S, Jurida L, Bartkuhn M, Wilhelm 3, Müller H, Beuerlein K, Karl N, Bhuju S, Ziebuhr J, Schmitz ML6, Kracht M (2017) The NF- κ B-dependent and-independent transcriptome and chromatin landscapes of human coronavirus 229E-infected cells. *PLoS Pathog* 13: e1006286. Doi: 10.1371/journal.ppat.1006286.
 26. [Loboda et al. 2019] Anna P Loboda, Surinder M Soond, Mauro Piacentini, Nickolai A Barlev (2019) Lysine-specific post-translational modifications of proteins in the life cycle of viruses. *Cell Cycle* 18(17): 1995-2005. Doi.org/10.1080/15384101.2019.1639305.
 27. Uhler and Shivashankar (2020) Coronaviruses and Its Interplay with Ageing. *Nat Rev Mol Cell Biol* 5: 247-248. Doi: 10.1038/s41580-020-0242-z.
 28. To Sing Fung, Ding Xiang Liu Review (2018) Post-translational modifications of coronavirus proteins: roles and function. *Future Virology* 13: 6. Doi.org/10.2217/fvl-2018-0008.
 29. Lin Chen, Oliver T. Keppler and Christian Schölz (2018) Post-translational Modification-Based Regulation of HIV Replication. *Front. Microbiol* 9: 2131. Doi.org/10.3389/fmicb.2018.02131.
 30. Mallagaray A, Creutzmacher R, Dülfer J, Mayer PHO, Grimm LL, Orduña JM, Trabjerg E, Stehle T, Rand KD, Blaum BS, Uetrecht C, Peters T (2019) A post-translational modification of human Norovirus capsid protein attenuates glycan binding. *Nature Communications* 10. Article number: 1320 (2019). Doi.org/10.1038/s41467-019-09251-5.
 31. [Liu et al. 2014] T Liu, U Sae-Ueng, D Li, G C Lander, X Zuo, B Jonsson, D Rau, I Shefer, A Evilevitch (2014) Solid-to-fluid-like DNA transition in viruses facilitates infection. *PNAS. Proceedings of the National Academy of Sciences* 111(41): 14675-14680. Doi.org/10.1073/pnas.1321637111.
 32. Udom Sae-Ueng, Dong Li, Xiaobing Zuo, Jamie B Huffman, Fred L Homa, Donald Rau, Alex Evilevitch (2014) Solid-to-fluid DNA transition inside HSV-1 capsid close to the temperature of infection. *Nature Chemical Biology* 10: 10: 861 Doi: 10.1038/nchembio.1628.
 33. Edoardo Salladini, Claire Debarnot, Vincent Delaunz, Maria Grazia Murrari, Priscila Sutto-Ortiz, Silvia Spinelli, Roberta Pierattelli, Christophe Bignon, Sonia Longhi (2018) Phase transition and amyloid formation by a viral protein as an additional molecular mechanism of virus-induced cell toxicity. *BioRxiv*. 2018. Posted December 14, 2018. Doi.org/10.1101/497024.
 34. Heinrich BS, Maliga Z, Stein DA, Hyman AA, and Whelan SPJ (2018) Peter Palese, Editor. Phase Transitions Drive the Formation of Vesicular Stomatitis Virus Replication Compartments. *MBio* 9(5): e02290-17. Doi: 10.1128/mBio.02290-17.
 35. Ricard Sole, Josep Sardany'es, Santiago F. Elena (2020) Phase Transitions in Virology. Preprints (www.preprints.org) Posted: 18 February 2020. Doi:10.20944/preprints202002.0261.v1.
 36. [Guseva et al. 2020] Serafima Guseva, Sigrid Milles, Malene Ringkjøbing, Jensen, Nicola Salvi, Jean-Philippe Kleman, Damien Maurin, Rob W. H. Ruigrok, Martin Blackledge (2020) Measles virus nucleo- and phosphoproteins form liquid-like phase-separated compartments that promote nucleocapsid assembly. *Science Advances* 6(14): 7095. Doi: 10.1126/sciadv.aaz7095. Phase Transitions in Membrane-Less-Oeganelles.
 37. Shi Y, Wang Y, Shao C, Huang J, Gan J, Huang X, Bucci E, Piacentini, M, Ippolito G, Melino G (2020) COVID-19 infection: the perspectives on immune responses *Cell Death and Differentiation* 27: 1451-1454. Doi.org/10.1038/s41418-020-0530-3.
 38. Grohmann U, and Bronte V (2010) Control of Immune Response by Amino Acid Metabolism *Immunol Rev* 236: 43-264. Doi: 10.1111/j.1600-065X.2010.00915.x.
 39. McGaha TL, Huang, L, Lemos H, Metz R, Mautino, M, Prendergast GC, Mellor AL (2012) Amino acid catabolism: a pivotal regulator of innate and adaptive immunity. *Immunol Rev* 249(1): 135-157. Doi: 10.1111/j.1600-065X.2012.01149.x
 40. Jumpei Sasabe, Masataka Suzuki. Review (2018) Emerging Role of D-Amino Acid Metabolism in the Innate Defense. *Front. Microbiology* 9: 933. Doi: 10.3389/fmicb.2018.00933.
 41. Alena Aliashkevich, Laura Alvarez and Felipe Cava. Review (2018) New Insights into the Mechanisms and Biological Roles of D-Amino Acids in Complex Eco-Systems. *Front. Microbiol.* 2018. Doi.org/10.3389/fmicb.2018.00683.
 42. Bastings JJAJ, Hans M. van Eijk, Steven W. Olde Damink, Sander S. Rensen. Review (2019) D-amino Acids in Health and Disease: A Focus on Cancer. *Nutrients* 11(9): 2205. Doi.org/10.3390/nu11092205.
 43. Silke Meiners, Oliver Eickelberg (2015) Melanie Königshoff Hallmarks of the ageing lung. *European Respiratory Journal* 45: 807-827; Doi: 10.1183/09031936.00186914.
 44. Matsui R, Goldstein RH, Mihal K, Brody JS, Steele MP, Fine A (1994) Type I collagen formation in rat type II alveolar cells immortalised by viral gene products. *Thorax* 49: 201-206.
 45. George PM, Wells AU, Jenkins RG (2020) Pulmonary fibrosis and COVID-19: the potential role for antifibrotic therapy. *The Lancet. Respiratory Medicine* Pg. 1-9. Doi.org/10.1016/S2213-2600(20)30225-3.

46. Peng TY, Lee KR, Tarn WY (2008) Phosphorylation of the arginine/serine dipeptide-rich motif of the severe acute respiratory syndrome coronavirus nucleocapsid protein modulates its multimerization, translation inhibitory activity and cellular localization. *FEBS J* 275(16): 4152-4163. Doi: 10.1111/j.1742-4658.2008.06564.x
47. Yingchun Zeng, Linbai Ye, Shengli Zhu, Hong Zheng, Peng Zhao, Weijia Cai, Liya Su, Yinglong She, Zhenghui Wu (2008) The nucleocapsid protein of SARS-associated coronavirus inhibits B23 phosphorylation. *Biochem Biophys Res Commun* 369(2): 287-291. Doi: 10.1016/j.bbrc.2008.01.096.
48. Spencer KA, Dee M, Paul Britton P, Hiscox JA (2008) Role of phosphorylation clusters in the biology of the coronavirus infectious bronchitis virus nucleocapsid protein. *Virology* 370(2): 373-381. Doi.org/10.1016/j.virol.2007.08.016.
49. Takamatsu Y, Krähling V, Kolesnikova L, Halwe S, Lier C, Baumeister S, Noda T, Biedenkopf N, Becker S (2020) Serine-arginine protein kinase 1 regulates Ebola virus transcription. *MBio*. 2020. 11:e02565-19. Doi.org/10.1128/mBio.02565-19.virus proteomics host-microbe/virus biology
50. Suh H, Ficarro SB, Kang UB, Chun Y, Marto JA, Buratowski S (2016) Direct Analysis of Phosphorylation Sites on the Rpb1 C-Terminal Domain of RNA Polymerase II. *Molecular Cell* 61: 297-304. Doi.org/10.1016/j.molcel.2015.12.021.
51. Fraser KA, Stephen A Rice SA (2005) Herpes Simplex Virus Type 1 Infection Leads to Loss of serine-2 Phosphorylation on the Carboxyl-Terminal Domain of RNA Polymerase II. *J. of Virology* 79(17): 11323-11334. Doi: 10.1128/JVI.79.17.11323-11334.2005.
52. Czudnochowski N, Bösken CA, Geyer M (2012) Serine-7 but not serine-5 phosphorylation primes RNA polymerase II CTD for P-TEFb recognition. *Nature Communications* 3. Article number: 842 (2012). Doi.org/10.1038/ncomms1846.
53. Cheng J, Tao J, Li B, Shi Y, Liu H (2019) The tyrosine 73 and serine 83 dephosphorylation of H1N1 swine influenza virus NS1 protein attenuates virus replication and induces high levels of beta interferon. *Virology Journal* 6. Article number: 152 (2019).
54. Kido H, Takahashi E, Kimoto T (2019) Role of host trypsin-type serine proteases and influenza virus-cytokine-trypsin cycle in influenza viral pathogenesis. Pathogenesis-based therapeutic options. *Biochim* 166: 203-213. Doi.org/10.1016/j.biochi.2019.09.006.
55. Shahiduzzaman M, Coombs K (2012) Activity based protein profiling to detect serine hydrolase alterations in virus infected cells. *Frontiers in Microbiology*. 2012. 3. Article 308 | 1. Doi: 10.3389/fmicb.2012.00308.
56. Heald-Sargent T, Gallagher T (2012) Ready, Set, Fuse! The Coronavirus Spike Protein and Acquisition of Fusion Competence. *Viruses* 4(4): 557-580. Doi.org/10.3390/v4040557.
57. Mo Liu, Chunfang Gu, Jianguo Wu, Ying Zhu (2006) Amino acids 1 to 422 of the spike protein of SARS associated coronavirus are required for induction of cyclooxygenase-2. *Virus Genes*. 2006. 33(3): 309-317. Doi: 10.1007/s11262-005-0070-4.
58. Shulla A, Heald-Sargent T, Subramanya G, Zhao J, Perlman P, Gallagher T (2011) A Transmembrane Serine Protease Is Linked to the Severe Acute Respiratory Syndrome Coronavirus Receptor and Activates Virus Entry. *J. of Virology* 85(2): 873-882. Doi: 10.1128/JVI.02062-10.
59. Jacob T, Van den Broeke C, Herman W, Favoreel HW (2011) Viral Serine/Threonine Protein Kinases. *J. of Virology* 85(3): 1158-1173. Doi:10.1128/JVI.01369-10.
60. Takahashi O, Kirikoshi R, Manabe N (2017) Racemization of Serine Residues Catalyzed by Dihydrogen Phosphate Ion: A Computational Study. *Catalysts* 7: 363. Doi:10.3390/catal7120363.
61. Ahn SH, Minkyu Kim, Stephen Buratowski (2004) Phosphorylation of Serine 2 Within the RNA Polymerase II C-terminal Domain Couples Transcription and 3' End Processing. *Mol Cell* 13(1): 67-76. Doi: 10.1016/s1097-2765(03)00492-1. Doi: 10.1016/s1097-2765(03)00492-1.
62. Macek B, Gnad F, Soufi B, Kumar C, Olsen JV, Mijakovic I, Mann M (2008) Phosphoproteome Analysis of *E. coli* Reveals Evolutionary Conservation of Bacterial Ser/Thr/Tyr Phosphorylation. *Molecular & Cellular Proteomics* 7(2): 299-307.
63. Miao B, Xiao Q, Chen W, Li Y, Zhen Wang Z (2018) Evaluation of functionality for serine and threonine phosphorylation with different evolutionary ages in human and mouse. *BMC Genomics*. 2018. 19. Article number: 431. Doi.org/10.1186/s12864-018-4661-6.
64. Barki-Harrington L, Elkobi A, Tzabary T, Rosenblum K (2009) *The Journal of Neuroscience* 29(29): 9219-9226. Doi.org/10.1523/JNEUROSCI.5667-08.2009.
65. Cairns DM, Rouleau N, Parker RN, Walsh KG, Gehrke L, Kaplan DL (2020) A 3D human brain-like tissue model of herpes-induced Alzheimer's disease. *Science Advances* 6:19: eaay8828. Doi: 10.1126/sciadv.aay8828. 63. 55.
66. Ha S, Kim I, Takata T, Kinouchi T, Isoyama M, Suzuki M, Noriko Fujii N (2017) Identification of D-amino acid-containing peptides in human serum. *PLOS-ONE* 2(12): e0189972. Doi.org/10.1371/journal.pone.0189972.
67. Hesaka A, Sakai S, Hamase K, Ikeda T, Matsui R, Mita M, Horio M, Isaka Y, Kimura T (2019) D-Serine reflects kidney function and diseases. *Scientific Reports* 9. Article number: 5104 (2019). Doi.org/10.1038/s41598-019-41608-0.
68. Kimura T, Hesaka A, Isaka Y (2020) D-Amino Acids and Kidney Diseases. *Clin Exp Nephrol* 24(5): 404-410. Doi: 10.1007/s10157-020-01862-3.
69. Xie P, Ma W, Tang H, Liu D (2020) Severe COVID-19: A Review of Recent Progress with a Look Toward the Future. *Mini Review. Front. Public Health*. 2020. Doi.org/10.3389/fpubh.2020.00189.
70. Mauricio G. Mateu (2013) *Structure and Physics of Viruses: An Integrated Textbook*. Pg. 59. 2.3 Viral Capsid Symmetry. Springer. 2013.
71. Sanjay D, Fangming X, Robijn B, William K, Joseph R (2016) Chirality of Viral Capsids. *APS March Meeting*.

2016. Abstract id. B37.002.
72. Konevtsova OV, Rochal SB, Lorman VL (2012) Chiral Quasicrystalline Order and Dodecahedral Geometry in Exceptional Families of Viruses. *Phys. Rev. Lett* 108: 038102. Doi.org/10.1103/PhysRevLett.108.038102.
 73. Li Y, Li H, Fan R, Wen B, Zhang J, Cao X, Wang C, Song Z, Shuoichi Li S, Li X, Lv X, Qu X, Huang R, Liu W (2016) Coronavirus Infections in the Central Nervous System and Respiratory Tract Show Distinct Features in Hospitalized Children. *Intervirology* 59(3): 163–169. Doi: 10.1159/000453066.
 74. Fang Li (2016) Structure, Function, Evolution of Coronavirus Spike Proteins. *Annu Rev Virol* 3(1): 237–261. Doi: 10.1146/annurev-virology-110615-042301.
 75. Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Velesler D (2020) Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell*. 2020. 181(2): 281-292. Doi.org/10.1016/j.cell.2020.02.058
 76. Verdiá-Báguena C, Nieto-Torres JL, Alcaraz A, DeDiego ML, Enjuanes L, Aguilera VM (2013) Analysis of SARS-CoV E protein ion channel activity by tuning the protein and lipid charge. *Biochim Biophys Acta* 1828(9): 026–2031. Doi: 10.1016/j.bbame.2013.05.008.

Copyright: ©2020 Victor V. Dyakin. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.